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CHIRAL SYNTHESIS OF D- AND L-3,3-DIPHENYLALANINE (DIP), UNUSUAL α-AMINO ACIDS FOR PEPTIDES OF BIOLOGICAL INTEREST

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Abstract: The asymmetric syntheses of D-(R)-(-)- and L-(S)-(+)-3,3-diphenylalanine (Dip) are described.

Currently, there is great interest in the design and synthesis of unusual α -amino acids which may provide specific conformational restraint, and also biostability to degradation by peptidases to small peptides/peptidomimetics as potential therapeutic agents.¹ In this regard, we undertook the preparation of multigram quantities of D- and L-3,3-diphenylalanine (Dip). Although these compounds have been incorporated into peptides as a racemate, and then separated as diastereoisomers,^{2,3} attempts at resolution by the action of enzymes such as hog kidney acylase, or carboxypeptidase on N-acetyl Dip are reported to be unsuccessful.² Our experiments at resolution using other commercial acylases also failed. However, we were able to obtain large quantities of D- and L-Dip by conventional resolution method using cinchona alkaloids.⁴ The absolute configuration of the resolved isomers (1a), ($[\alpha]_D^{23} = -64.5^\circ$ (c=1.0, MeOH)) as D-Dip hydrochloride and (1b), ($[\alpha]_D^{23} = +63.9^\circ$ (c=1.0, MeOH)) as L-Dip hydrochloride were tentatively assigned on the basis of the order of elution on a Diacel Crownpack HPLC column. ^{5,6,7,8} This assignment of absolute configuration could not be confirmed by a single crystal x-ray



analysis,⁹ since a single crystal of the compound or a derivative thereof could not be obtained. In order to confirm the assigned configurations, we undertook the chiral synthesis of D- and L-Dip using Evans methodology,¹⁰ which confirmed our original assignment of configuration. A recent report by Chassaing *et. al.*,¹¹ on the asymmetric synthesis of these same amino acids via an alternative sultam derived chiral auxiliary methodology (4 steps, 46% overall yield, 95% d.e., long reaction times),



The conversion of 3,3-diphenylpropionic acid (2) (1.1 equiv.), to the mixed anhydride (3) was accomplished by the treatment of the acid (2) with DIPEA (1.2 equiv.) and pivalovl chloride (1.05 equiv.) in THF at -78°C. The milky mixture was warmed to 0°C over 60 min. The solid was filtered and the filtrate was added to the lithiated oxazolidinone solution at -78°C, which was prepared by the addition of n-BuLi (1.0 equiv.) to the solution of (4R,5S)-4-methyl-5-phenyl-2-oxazolidinone (4a), (1.0 equiv.) in dry THF. The yellow solution was warmed to 0°C, and quenched with water. The residue, after standard work-up. was flash chromatographed (silica gel, hexane: AcOEt/4 : 1) to yield the acyloxazolidinone (5a).¹² 82% yield, mp 106°C, $[\alpha]_{D}^{23} = -13.5^{\circ}$ (c=1.0, MeOH). Deprotonation of (5a) by the addition of KHMDS (1.05 equiv.) in THF at -78°C, followed by the addition of a solution of trisyl azide (6)¹⁰(1.25 equiv.) in THF, rapidly quenching with AcOH (4.6 equiv.) after 2 min at -78°C, and warming quickly to 30°C gave a light yellow solution, which was diluted with CH₂Cl₂, washed with brine, NaHCO₂ (sat.), and dried (MgSO₄). The concentrated yellow oil was flash chromatographed (silica gel, hexane: AcOEt/8 : 1) to give the azido oxazolidinone (7a), 94% yield, mp 90 - 91°C, $[\alpha]_D^{23} = -206.2^\circ$ (c=1.0, MeOH). HPLC¹³ showed a single diastereoisomer indicating complete stereocontrol during azide entry onto the potassium enolate template of (5a). The azide (7a) in 4 : 1 THF-H₂O at 0°C, was hydrolyzed with a solution of LiOH-H₂O (2.0 equiv.) in H₂O₂ (5.0 equiv., 30% aqueous). After 1 hr., aqueous Na₂SO₃ (1 M) was added. THF was removed in vacuo, and the aqueous was acidified to pH 1 with 6 N HCl at 0°C. The product was extracted with CH₂Cl₂, and dried (MgSO₄). The azido acid (8a) was obtained by flash chromatography (silica gel, hexane:AcOEt:AcOH/50 : 50 : 2), 99% yield, mp 108 - 109.5°C, $[\alpha]_D^{23} = -54.2^{\circ}$ (c=1.0, MeOH).

The chiral auxiliary (4a) was recovered (96%, mp 120 - 121°C). The azido acid (8a) in THF-HCl (1 N), (4 : 1), was hydrogenated at 50 psi, over 10% Pd-C, for 20 hrs. The solvent was removed, and the residue was recrystallized from 2N HCl

Scheme I

to give a white solid, as D-(R)-(·)-Dip hydrochloride (1a), 77% yield, mp 248 - 253°C (dec.), $[\alpha]_D^{23} = -63.7^\circ$ (c=1.0, MeOH),-¹⁴ Chiral HPLC¹⁵ indicated a >99% enantiomeric purity. Further treatment of (1a) with (Boc)₂O (2.0 equiv.) and DIPEA (2.5 equiv.) in methanol gave N-Boc-D-Dip (9a), 81% yield, mp 153 - 154°C (dec.), $[\alpha]_D^{23} = -35.7^\circ$ (c=1.0 MeOH).

The L-(S)-(+)-DIP hydrochloride (1b) (mp 247 - 250°C (dec.), $[\alpha]_D^{23} = +63.8^\circ$ (c=1.0, MeOH)) and N-Boc-L-Dip (9b)¹⁶ (mp 152 - 153°C (dec.), $[\alpha]_D^{23} = +32.2^\circ$ (c=1.0, MeOH)) were similarly synthesized using (4S,5R)-4-methyl-5-phenyl-2-oxazolidinone (4b)¹⁷ as the chiral auxiliary (Scheme II).

Scheme II



In conclusion, the synthetically useful D-(R)-(-)-Dip and L-(S)-(+)-Dip can be enantioselectively prepared in good yield using Evans methodology. The predictable stereochemistry of the α -amino acids obtained from this methodology confirmed the absolute configuration assigned to the enantiomers, obtained by conventional resolution, using fast and easily available chiral HPLC techniques.

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- 8. The absolute configuration of the resolved Dip was assigned by chiral HPLC with the same conditions as described in note
- 15, the isomer with t_R (retention time) = 80.76 min assigned as D-(-)-Dip and t_R = 89.75 min as L-(+)-Dip.
- 9. We thank Professor A. T. McPhail (Duke University) for this effort.

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11. a) Josien, H.; Martin, A.; Chassaing, G.; *Tetrahedron Lett.*, **1991**, *32*, 6547. b) The rotation and mp of D- and L-Dip we found are significantly higher than those reported by Chassaing. The enantiomeric purity of our D- and L-Dip is confirmed by chiral HPLC analysis (see note 8 and 15).

12. The acyloxazolidinone (**5a**) was also prepared using 3,3-diphenylpropionyl chloride, (mp 41 - 43°C), instead of the mixed anhydride which gave (**5a**) in 80% yield, mp 106°C, recrystallized from hexane-AcOEt (4 : 1).

13. Analytical reverse-phase HPLC was performed with a 4.6 x 250 mm, 5 μ M, C-18, Beckman Ultrasphere column, solvent A: 0.05 M NH₄H₂PO₄ in water; solvent B: acetonitrile, A:B/45:55, to give one peak, t_B = 11.73 min.

14. Spectral data for: (5a): IR(KBr): 1784.0, 1764.5, 1494.6, 1454.1, 1350.2 cm⁻¹. ¹H NMR (CDCl₂): δ 7.40-7.13 (m, 15H), 5.46 (d, J=8.76 Hz, 1H), 4.68 (t, J₁=J₂=9.39 Hz, 1H), 4.64-4.53 (m, 1H), 3.78 (d, J=8.61 Hz, 2H), 0.71 (d, J=7.92 Hz, 3H). ¹³C NMR(CDCl₂): δ 171.16, 153.00, 143.43, 143.34, 133.19, 128.71, 128.64, 128.52, 127.90, 127.78, 126.55, 126.50, 125.59, 78.89, 54.70, 46.77, 40.92, 14.29. MS(CI): m/z 386 (M⁺+1), 385 (M⁺), 222, 167 (base), 165, 77. Anal.: Calcd. for C25H23NO3: C, 77.90; H, 6.01; N, 3.63. Found: C, 77.86; H, 6.11; N, 3.63. (7a): IR(KBr): 2109.7, 1783.6. 1734.1 cm^{-1} . ¹H NMR(CD₃OD): δ 7.49-7.24 (m, 15H), 6.04 (d, J=13.26 Hz, 1H), 5.03 (d, J=8.55 Hz, 1H), 4.55 (d, J=13.32 Hz) Hz, 1H), 4.45-4.34 (m, 1H), 0.81 (d, J=7.98 Hz, 3H). ¹³C NMR(CD₂OD): δ 171.36, 153.97, 141.06, 134.37, 129.93, 129.90, 129.73, 129.63, 129.50, 128.66, 128.49, 128.29, 127.18, 126.89, 80.70, 62.38, 56.44, 54.96, 14.48. MS(FAB): m/z 427 (M⁺+1), 401 (base), 386. Anal.: Calcd. for C₂₅H₂₂N₄O₃: C, 70.41; H, 5.20; N, 13.14. Found: C, 70.06; H, 5.22; N, 12.79. (8a): IR(KBr): 3416.5-3303.9(br), 2121.9, 1739.4 cm⁻¹. ¹H NMR(CD₃OD): δ 7.41-7.14 (m, 10H), 4.65 (d, J=11.16 Hz, 1H), 4.50 (d, J=11.37 Hz, 1H). ¹³C NMR(CD₃OD): § 172.69, 141.86, 141.51, 129.75, 129.64, 129.49, 128.17, 66.62, 53.97. MS(CI): m/z 222 (M⁺.45), 196, 195, 194, 167 (base). Anal.: Calcd. for C₁ sH₁₃N₃O₅: C, 67.41; H, 4.90; N, 15.72. Found; C, 67.31; H, 4.90, N, 15.03. (1a): IR(KBr): 3508.9-3373.7(br), 3361.9-3268.0(br), 1757.5 cm⁻¹. ¹H NMR(CD₃OD): δ 7.52-7.17 (m, 10H), 4.85 (d, J=12.66 Hz, 1H), 4.43 (d, J=12.63 Hz, 1H). ¹³C NMR(CD₃OD): δ 171.04, 140.24, 139.65, 130.50, 129.81, 129.52, 129.41, 129.17, 128.70, 57.41, 54.84. MS(CI): m/z 242 (M⁺+1), 196, 167 (base). (9a): IR(KBr): 3386.6-3372.1(br), 1716.9, 1700.4 cm⁻¹. ¹H NMR(CD₃OD); δ 7.34-7.15 (m, 10H), 4.97 (d, J=13.29 Hz, 1H), 4.34 (d, J=13.20 Hz, 1H), 1.31 (s, 9H). ¹³C NMR(CD₃OD): δ 176.27, 155.53, 140.11, 139.40, 128.78, 128.71, 128.60, 128.26, 127.28, 127.12, 80.59, 56.60, 52.63, 28.20. MS(CI): m/z 342 (M⁺+1), 286, 242, 196, 167 (base). Anal: Calcd. for C20H23NO4: C, 70.36; H, 6.79; N, 4.10. Found: C, 70.31; H, 6.86; N, 3.94.

15. The chiral HPLC was performed with 4.0 x 150 mm chiral crown ether column, Daicel Crownpak CR(+) and Varian Star HPLC System, solvent A: $HCIO_4$ in H_2O , pH 1.0, solvent B: MeOH, A:B/86:14, flow rate 0.6 mL/min., to give (1a) $t_R = 80.86$ min., purity 99.9% by peak area normalization, (1b) $t_R = 86.30$ min., purity 99.3% by peak area normalization.

16. Spectral data for: (1b): IR(KBr): 3450-3321 (br), 3090.7-2830.6 (br), 1757.4 cm⁻¹. ¹H NMR(CD₃OD): δ 7.52-7.19 (m, 10H), 4.86 (d, J=12.54 Hz, 1H), 4.43 (d, J=12.54 Hz, 1H). ¹³C NMR(CD₃OD): δ 171.05, 140.25, 139.66, 130.52, 129.83, 129.55, 129.44, 129.17, 128.71, 57.44, 54.83. MS(CI): m/z 242 (M⁺+1), 196, 167 (base). (9b): IR(KBr): 3388.2(br), 3380.5(br), 1700.1, 1695.8 cm⁻¹. ¹H NMR(CD₃OD): δ 7.35-7.17 (m, 10H), 4.97 (d, J=10.92 Hz, 1H), 4.36 (d, J=10.68 Hz, 1H), 1.32 (s, 9H). ¹³C NMR(CDCl₃): δ 176.79, 155.55, 140.30, 139.60, 128.77, 128.60, 128.33, 127.21, 127.07, 80.55, 56.81, 52.73, 28.24. MS(FAB): m/z 342 (M⁺+1), 286, 242 (base). Anal: Calcd. for C₂₀H₂₃NO₄: C, 70.36; H, 6.79; N, 4.10. Found: C, 69.98; H, 6.95; N, 3.87.

17. Compounds (4b), (5b), (7b), (8b) are enantiomers of (4a), (5a), (7a), (8a) respectively.

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